## NOTE



## **Open Access**

# PLANT U-BOX PROTEIN 10 negatively regulates abscisic acid response in Arabidopsis

Jun Sung Seo<sup>1,2</sup>, Pingzhi Zhao<sup>1,3</sup>, Choonkyun Jung<sup>1,4\*</sup> and Nam-Hai Chua<sup>1,2\*</sup>

### Abstract

MYC2 is well known as a positive regulator for abscisic acid (ABA) signaling but whether PLANT U-BOX PROTEIN 10 (PUB10) is involved in ABA responses has not been reported. Here, we show that the E3 ubiquitin ligase PUB10 modulates ABA signaling in Arabidopsis. *PUB10ox (35S:PUB10-myc)* and *myc2* loss-of-function mutants were hyposensitive to ABA during germination, whereas *pub10* loss-of-function and *MYC2ox (35S:MYC2-GFP)* mutants were hypersensitive. In addition, *pub10* mutants showed hypersensitivity to high salt and osmotic stress during germination; by contrast, *PUB10ox* line displayed the opposite phenotype. ABA-induced expression of *KIN2 (Cold- and ABA-Inducible Protein)*, *RD22 (Responsive to Dehydration 22), ANAC019 (NAC Domain-Containing Protein 19)*, and *ANAC055 (NAC Domain-Containing Protein 55)* was enhanced in both *pub10* and *MYC2ox* plants. Taken together, *pub10* plants phenocopied *MYC2ox* plants, whereas *PUB10ox* plants phenocopied *myc2* in ABA response. Our results provide evidence that PUB10 negatively regulates ABA signaling in Arabidopsis.

Keywords: Abscisic acid, ANAC019, ANAC055, Arabidopsis, MYC2, PUB10

### Introduction

Abscisic acid (ABA) is a phytohormone present in all vascular plants and it participates in various developmental and physiological processes during the plant life cycle, including seed development, seed dormancy, germination, and abiotic stress responses [1]. The phosphorylation and dephosphorylation of protein are key post-translational modifications in ABA signal transduction [2]. In addition, regulation of protein stability via ubiquitination of key components of the ABA signaling pathways also plays an important role [3].

E3 ubiquitin ligases are responsible for the specificity of ubiquitination by recruiting appropriate target proteins [4]. Until now, only a limited number of Plant U-box (PUB) E3 ligases have been characterized as both positive and negative regulators of ABA signaling [5]. For instance, PUB12 and PUB13 were found to ubiquitinate

National University, Pyeongchang, Republic of Korea

Full list of author information is available at the end of the article



ance 1 (PYR1) [6]. Another U-box E3 ligase, Carboxyl terminus of the Hsc70-Interacting Protein (CHIP), monoubiquitinated PP2A subunits and enhances their activities under stress conditions [7]. In addition, PUB9, PUB18, and PUB19 are involved in ABA signaling [8, 9].

ABI1 in the presence of both ABA and Pyrabactin Resist-

MYC2 has been well characterized as a central transcriptional regulator in JA signaling [10, 11]. MYC2 was first isolated as a transcription factor that binds to the *RESPONSIVE DEHYDRATION 22 (RD22)* promoter and subsequently characterized as a positive regulator of ABA signaling through genetic analysis [12, 13]. Hence, MYC2 seems to integrate at least two (ABA and JA) different signaling pathways to coordinate growth and development in Arabidopsis.

Crosstalk between ABA and JA signaling pathways has been poorly examined so far. Previously, we showed that MYC2 directly interacts with PUB10, and that PUB10 modulates JA signaling by destabilizing MYC2 protein [14]. However, how PUB10 regulates ABA responses is mostly unknown. Here, we showed that PUB10 is a negative regulator in ABA signaling.

© The Author(s) 2019. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

<sup>\*</sup>Correspondence: jasmin@snu.ac.kr; bchcnh@nus.edu.sg

<sup>&</sup>lt;sup>2</sup> Temasek Life Sciences Laboratory, National University of Singapore, Singapore, Singapore

<sup>&</sup>lt;sup>4</sup> Graduate School of International Agricultural Technology and Crop Biotechnology, Institute/GreenBio Science and Technology, Seoul

### **Materials and methods**

### Plant materials and growth conditions

Arabidopsis thaliana Columbia ecotype (Col-0), myc2 (jin1-9, SALK\_017005), pub10 (SALK\_017111), MYC20x, and PUB100x lines [14] were grown on 0.5% agar medium containing Murashige and Skoog (MS) salts, 1% sucrose, and 0.5 g/L MES hydrate at 22 °C under 16 h white fluorescent light (100  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>)/8 h dark. T-DNA insertion lines were obtained from the SALK collection [15].

### **Germination test**

Seeds were spread onto MS agar plates with or without 2  $\mu$ M ABA, 150 mM NaCl, or 200 mM mannitol. After 4 days of stratification at 4 °C, seed germination was monitored from 0 to 7 days.

### Abscisic acid treatments

Ten-day-old Col-0, *pub10*, and *MYC20x* Arabidopsis seedlings grown on MS agar plates were transferred to liquid MS medium containing 10  $\mu$ M ABA. Treated seed-lings were collected at the indicated time points for real-time RT-PCR analysis.

### **Real-time RT-PCR analysis**

Total RNA was extracted from Arabidopsis seedlings treated with ABA using an RNeasy Plant Mini Kit (Qiagen) including DNase I treatment. Reverse transcription was performed using 2  $\mu$ g of each total RNA and oligo (dT)<sub>20</sub> primers by SuperScript III reverse transcriptase (Invitrogen). Real-time RT-PCR was performed using SYBR premix Ex Taq (Tli RNaseH plus) (TaKaRa) on the Bio-Rad CFX96 real-time system with gene-specific primers. Primer sequences are listed in Additional file 1: Table S1.

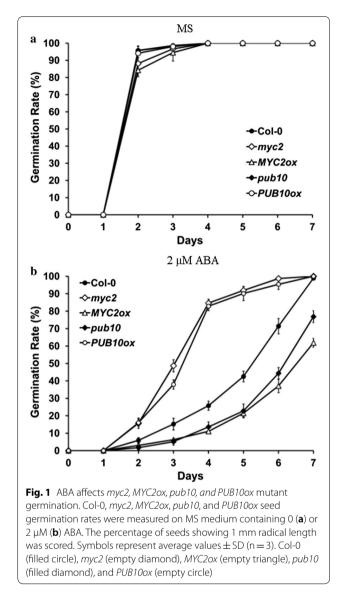
### Accession numbers

Sequence data can be found in the Arabidopsis Genome Initiative data library under the following accession numbers: ACT2 (AT3G18780), ANAC019 (AT1G52890), ANAC055 (AT3G15500), KIN2 (AT5G15970), MYC2 (AT1G32640), PUB10 (AT1G71020), RD22 (AT5G25610).

### **Results and discussion**

### PUB10 negatively regulates ABA response during seed germination

To examine PUB10 function in ABA responses, we analyzed the germination rates of *pub10*, *PUB100x* (35S:PUB10-myc), myc2 (jin1-9), and MYC20x (35S:MYC2-GFP) seeds on MS agar plates containing 2  $\mu$ M ABA. Germination rates of all genotypes in MS medium without ABA were almost the same as those of Col-0 (Fig. 1a). At 2  $\mu$ M ABA, the germination rates of *pub10* and *MYC20x* seeds were significantly reduced



compared to Col-0 seeds (Fig. 1b), but germination of *PUB10ox* and *myc2* seeds showed a remarkable ABAinsensitive phenotype (Fig. 1b). These results indicate that the altered germination rates observed in *PUB10* and *MYC2* mutants are dependent on ABA sensitivity, and that PUB10 and MYC2 act as positive and negative ABA response regulators, respectively. These opposing ABA sensitivities between *PUB10* and *MYC2* mutants are consistent with our previous observation that MYC2 protein is destabilized by the E3 ubiquitin ligase PUB10 [14].

# PUB10 positively regulates salt and osmotic stress tolerance during seed germination

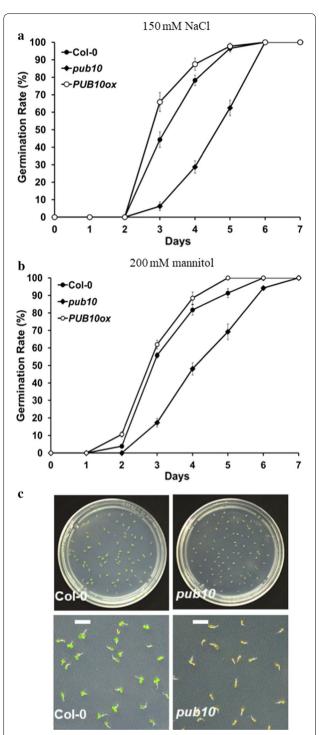
ABA plays a key role in abiotic stress responses including drought, salt, and osmotic stress [1]. To evaluate the

effect of PUB10 on salt and osmotic stress responses, we analyzed the seed germination rates of pub10 and PUB10ox on MS agar plates containing 150 mM NaCl or 200 mM mannitol. The germination rate of pub10 seeds was significantly decreased in high salt medium compared to Col-0 seeds, but PUB10ox seeds showed an increased germination rate compared to Col-0 seeds (Fig. 2a). The germination rates of pub10 and PUB10ox seeds in mannitol-containing medium were similar to those in high salt medium. PUB10ox and pub10 seeds showed tolerant and hypersensitive phenotypes to osmotic stress, respectively (Fig. 2b). Growth inhibition of *pub10* mutant seedlings was stronger than that of Col-0 seedlings (Fig. 2c). Taken together, these results indicated that PUB10 acts as a positive regulator for salt and osmotic stress tolerance.

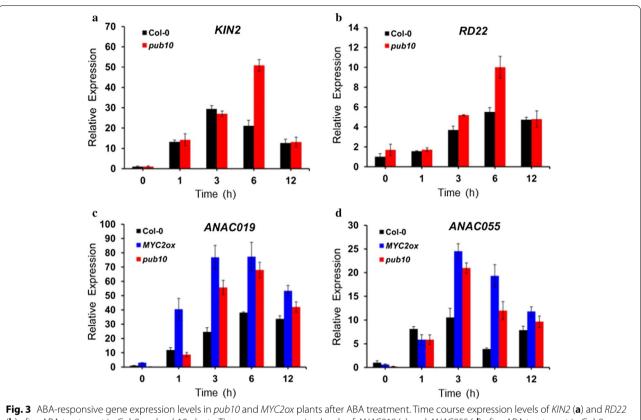
# PUB10 negatively regulates ABA-responsive gene expression

To investigate how PUB10 regulates gene expression in response to ABA, we performed a time-course analysis for ABA-responsive gene expression in pub10 and MYC2ox plants after ABA treatment. To determine the role of PUB10 in the MYC2-dependent ABA signaling pathway, we monitored the expression of KIN2, RD22, ANAC019, and ANAC055 genes previously reported as ABA-responsive genes regulated by MYC2 [12, 15, 16]. KIN2 and RD22 expression were enhanced more than twofold in pub10 mutants compared to Col-0 6 h after ABA treatment (Fig. 3a, b). A previous report showed that transcription levels of KIN2 and RD22 were enhanced in MYC2ox and reduced in myc2 after ABA treatment [12]. Therefore, enhanced expression of KIN2 and *RD22* in *pub10* could be caused by increased MYC2 protein level. Transcription levels of two NAC transcription factors, ANAC019 and ANAC055, were induced by ABA, dehydration, and salt, and are also known as direct MYC2 targets [15-18]. ANAC019 and ANAC015 expression were higher in *pub10* and *MYC2ox* plants than those in Col-0 plant at 3 to 6 h after ABA treatment (Fig. 3c, d). Similar expression patterns in *pub10* and *MYC2ox* after ABA treatment support the notion that PUB10 negatively regulates target gene expression by destabilizing MYC2 protein. Overall, our results indicate that PUB10 negatively regulates ABA or salt responses by modulating MYC2 protein levels.

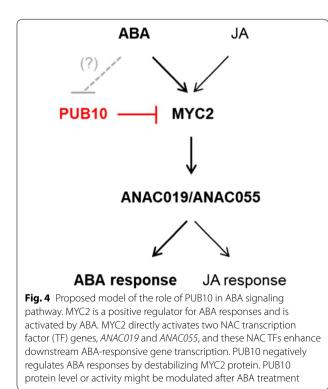
Although MYC2 has been first identified as a positive regulator of ABA responses [11, 12], MYC2 function in ABA signaling is almost unknown. Rather, MYC2 has been intensively studied as a key regulator in the JA signaling pathway [10]. MYC2 directly binds to the promoters of two NAC transcription factors, ANAC019 and ANAC055 and activates their transcription levels. These



**Fig. 2** *pub10* and *PUB10ox* germination in the presence of high salt and osmotic stress. Col-0, *pub10*, and *PUB10ox* seed germination rate on MS medium containing 150 mM NaCl (**a**) and 200 mM mannitol (**b**). Early post-germinative growth of Col-0 and *pub10* mutants 5 days after germination on 150 mM NaCl medium (**c**). The percentage of seeds showing 1 mm radical length was scored after 4 days of stratification at 4 °C. Seed germination was monitored from 0 to 7 days. Symbols represent average values  $\pm$  SD (n = 3). Col-0 (filled circle), *pub10* (empty triangle), and *PUB10ox* (empty circle). Scale bar = 10 mm



(b) after ABA reatment in Col-0 and *pub10* plants. Time course expression levels of *ANAC019* (c) and *ANAC055* (d) after ABA treatment in Col-0, *MYC20x*, and *pub10* plants. **a**-d transcript levels were normalized to *ACT2* expression levels, and bars represent average  $\pm$  SD (n = 3 independent seedling pools)



two NAC factors enhance the downstream transcription JA- and ABA-responsive genes, and also play key roles in the crosstalk between JA and ABA signaling (Fig. 4) [10, 15]. We suggest that PUB10 acts as a negative regulator of ABA signaling through MYC2; further study on the regulation of PUB10 protein by ABA is necessary to understand how PUB10 participates in the fine tuning of ABA signaling and JA crosstalk. Our results provide new insights for increasing ABA-mediated abiotic stress tolerance in plants.

### **Additional file**

Additional file 1: Table S1. Primers used in this study.

### Acknowledgements

We thank Bobby Williams, Cheng Lu, and Bongsoo Park for their helpful suggestions.

### Authors' contributions

CJ, and NHC conceived the experiments. JSS, PZ, and CJ conducted the experiments. JSS, CJ, and NHC analyzed the results. JSS, CJ, and NHC wrote the manuscript. All authors read and approved the final manuscript.

#### Funding

This work was supported in part by a grant from DuPont Company. C.J. was supported in part by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (NRF-2016R1D1A1B03932070) and a grant from the Next-Generation BioGreen 21 Program (PJ013399) of Rural Development Administration, Republic of Korea.

#### Availability of data and materials

Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

### **Competing interests**

The authors declare that they have no competing interests.

### Author details

<sup>1</sup> Laboratory of Plant Molecular Biology, Rockefeller University, New York, USA. <sup>2</sup> Temasek Life Sciences Laboratory, National University of Singapore, Singapore, Singapore. <sup>3</sup> State Key Laboratory of Plant Genomics, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China. <sup>4</sup> Graduate School of International Agricultural Technology and Crop Biotechnology, Institute/ GreenBio Science and Technology, Seoul National University, Pyeongchang, Republic of Korea.

### Received: 20 May 2019 Accepted: 15 July 2019 Published online: 27 July 2019

#### References

- 1. Finkelstein R (2013) Abscisic acid synthesis and response. Arabidopsis Book. 11:e0166
- Raghavendra AS, Gonugunta VK, Christmann A, Grill E (2010) ABA perception and signaling. Trends Plant Sci 15:395–401
- Lopez-Molina L, Mongrand S, Kinoshita N, Chua NH (2003) AFP is a novel negative regulator of ABA signaling that promotes ABI5 protein degradation. Genes Dev 17:410–418
- Vierstra RD (2009) The ubiquitin-26S proteasome system at the nexus of plant biology. Nat Rev Mol Cell Biol 10:385–397
- Yee D, Goring DR (2009) The diversity of plant U-box E3 ubiquitin ligases: from upstream activators to downstream target substrates. J Exp Bot 60:1109–1121
- Kong L, Cheng J, Zhu Y, Ding Y, Meng J, Chen Z et al (2015) Degradation of the ABA co-receptor ABI1 by PUB12/13 U-box E3 ligases. Nature Commun. 6:8630

- Luo JH, Shen GX, Yan JQ, He CX, Zhang H (2006) AtCHIP functions as an E3 ubiquitin ligase of protein phosphatase 2A subunits and alters plant response to abscisic acid treatment. Plant J. 46:649–657
- Samuel MA, Mudgil Y, Salt JN, Delmas F, Ramachandran S, Chilelli A et al (2008) Interactions between the S-domain receptor kinases and AtPUB-ARM E3 ubiquitin ligases suggest a conserved signaling pathway in Arabidopsis. Plant Physiol 147:2084–2095
- Bergler J, Hoth S (2011) Plant U-box armadillo repeat proteins AtPUB18 and AtPUB19 are involved in salt inhibition of germination in Arabidopsis. Plant Biol. 13:725–730
- 10. Kazan K, Manners JM (2013) MYC2: the master in action. Mol Plant. 6:686–703
- Lorenzo O, Chico JM, Sánchez-Serrano JJ, Solano R (2004) JASMONATE-INSENSITIVE1 encodes a MYC transcription factor essential to discriminate between different jasmonate-regulated defense responses in Arabidopsis. Plant Cell. 16:1938–1950
- Abe H, Yamaguchi-Shinozaki K, Urao T, Iwasaki T, Hosokawa D, Shinozaki K (1997) Role of Arabidopsis MYC and MYB homologs in drought- and abscisic acid-regulated gene expression. Plant Cell. 9:1859–1868
- Abe H, Urao T, Ito T, Seki M, Shinozaki K, Yamaguchi-Shinozaki K (2003) Arabidopsis AtMYC2 (bHLH) and AtMYB2 (MYB) function as transcriptional activators in abscisic acid signaling. Plant Cell. 15:63–78
- Jung C, Zhao P, Seo JS, Mitsuda N, Deng S, Chua NH (2015) PLANT U-BOX PROTEIN10 regulates MYC2 stability in Arabidopsis. Plant Cell. 27:2016–2031
- Alonso JM, Stepanova AN, Leisse TJ, Kim CJ, Chen H, Shinn P et al (2003) Genome-wide insertional mutagenesis of *Arabidopsis thaliana*. Science 301:653–657
- Jiang H, Li H, Bu Q, Li C (2009) The RHA2a-interacting proteins ANAC019 and ANAC055 may play a dual role in regulating ABA response and jasmonate response. Plant Signal Behav. 4:464–466
- Tran LS, Nakashima K, Sakuma Y, Simpson SD, Fujita Y, Maruyama K et al (2004) Isolation and functional analysis of Arabidopsis stress-inducible NAC transcription factors that bind to a drought-responsive cis-element in the early responsive to dehydration stress 1 promoter. Plant Cell. 16:2481–2498
- Gimenez-Ibanez S, Boter M, Ortigosa A, García-Casado G, Chini A, Lewsey MG et al (2017) JAZ2 controls stomata dynamics during bacterial invasion. New Phytol 213:1378–1392

### **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

# Submit your manuscript to a SpringerOpen<sup>®</sup> journal and benefit from:

- Convenient online submission
- ► Rigorous peer review
- Open access: articles freely available online
- ► High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at > springeropen.com