

Effects of Chlorine Concentrations and Washing Conditions on the Reduction of Microbiological Contamination in Lettuce

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Received October 27, 2008; Accepted December 25, 2008

We established optimum washing and sanitizing conditions for fresh-cut lettuce using sodium hypochlorite to reduce microbial hazards. Reduction of microbial hazards, including total aerobic bacteria, *Escherichia coli*, and *Staphylococcus aureus* at different sodium hypochlorite concentrations (0-500 ppm), immersion times (0-20 min), temperatures (0-30°C), and washing times (0-4 min) was evaluated. The optimum washing and sanitizing conditions using sodium hypochlorite for lettuce were determined as immersion at more than 50 ppm for 1 min at 20°C and washing twice for 30 sec after dipping. Application of these optimum conditions will improve safety and added value of lettuce as a fresh-cut-food without detrimental effects on sensory characteristics.

Key words: *fresh-cut-food, lettuce, sodium hypochlorite, washing optimum conditions*

Food consumption in Korea is changing toward convenience of use, better appearance, increased nutritional value, and better sanitation [Beuchat and Golden, 1985; Sakai S, 1995; Bae, 2001]. In Korea, like the USA and Europe, there is an increase in the amount and variety of vegetables that can meet the demands of consumers for freshness and convenience [Gardial *et al.*, 1994; Marshall *et al.*, 1995; Peter *et al.*, 2004]. A reflection of these consumer demands is a new product category of 'fresh-cut-food', which is a raw farm product subjected to simple processes such as cleaning, peeling, cutting, and mincing or addition of food additives. Fresh-cut-foods include salads and fresh vegetables that can be consumed without preparation. Fresh-cut-foods are affected by changes in the plant respiration rate and

enzyme activity due to tissue scarring that occurs during cutting and peeling. These types of foods are subject to contamination by pathogenic microorganisms during processing and distribution [Kumimoto, 1997]. In addition, since most fresh-cut-foods are used raw, like salads, bacterial safety is a concern [Kim *et al.*, 2002]. In the USA, many cases of food-borne disease caused by fresh-cut-foods have been reported and *E. coli* O157:H7 was first confirmed in 1982 as a human host bacterium that causes food-borne illness in the USA [Riley *et al.*, 1983]. Food-borne outbreaks of *Staphylococcus aureus* (*S. aureus*) occur in foods like lettuce and cucumber [Bergdoll, 1979]. Specific sources of food-borne illness due to fresh-cut-foods since 1990 are ranked by category from highest to lowest as salads, followed by fruits, germinating vegetables, cabbage, and carrots. By pathogen, from highest to lowest for cause of illness is *Salmonella*, *E. coli* O157:H7, *Shigella*, and *B. cereus*, and the frequency of occurrence is steadily increasing. A case of *Salmonella* food-borne occurrence in tomato was reported in 2008 [USFDA, 2008].

To prevent food-borne disease outbreaks, fresh-cut-foods should be protected from food-borne contamination

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Abbreviations: CFU, colony-forming unit; *E. coli*, *Escherichia coli*; *S. aureus*, *Staphylococcus aureus*

during processing and storage [Manvell and Ackland, 1986]. Due to their characteristics, fresh-cut-foods should not be sterilized by heating, so chemical sterilization using halogen compounds, oxidizers, alcohols, and physical methods using radiation or UV irradiation are being used [Jung *et al.*, 1996; No, 2003]. Among these, chemical methods are most widely used for convenience and cost effectiveness. In the USA, issues of safety regarding pathogenic microorganisms on fresh agricultural products are recognized, so most fresh agricultural products are processed with a no-rinse sterilizer. Compounds used with the approval of the U.S FDA as sterilizers for food surfaces include chlorine, chlorine dioxide, iodophors, quaternary ammonium compounds, and acid anions [Choi and Lee, 2008]. Among these, chlorines are most widely used for control of microorganisms in the food industry in Korea [Kim, 2001; Kim *et al.*, 2002]. Sodium hypochlorite, an inorganic chlorine, is the most widely used chemical due to convenience. There is not much research relating to reduction of microorganisms in leafy vegetables [Takeuchi and Frank, 2000], so sterilizers are used indiscriminately without scientific verification of effective concentrations, optimal washing conditions, and sterilizing effects.

The purpose of this study was to establish optimum washing and sanitizing conditions using a sodium hypochlorite solution treatment for fresh-cut lettuce to reduce microbial contamination.

Materials and Methods

Samples and inoculation. Lettuce was purchased from a local wholesale market in Ansong, Korea. Lettuce samples were inoculated with a suspension of *Escherichia coli* (*E. coli*) ATCC 10536 and *S. aureus* ATCC 6538 (0.1 mL in 100 g of lettuce) to a final concentration of 10^3 - 10^4 CFU/g, and dried on clean bench for sampling. Chlorine solutions were prepared using sodium hypochlorite (4% w/v, Yuhan Corporation, Seoul, Korea). These solutions were made just before the experiment and then used. To determine an optimum sodium hypochlorite concentration, lettuce (10 g) was immersed in a sterile beaker containing 200 mL of sodium hypochlorite (0-500 ppm) for 5 min following the School Foodservice Health Control Guidebook (for vegetables) and washed in 200 mL of tap water for 1 min. To determine an optimum immersion time in the sodium hypochlorite solution, lettuce (10 g) was immersed in a sterile beaker containing 200 mL of sodium hypochlorite (200 ppm) for 0-20 min and washed in 200 mL of tap water for 1 min. To determine optimum washing times, lettuce (10 g) was immersed in a sterile beaker containing 200 mL of

sodium hypochlorite solution (200 ppm) for 5 min and washed in 200 mL of tap water from one to eight times for 30 sec. To determine an optimum sodium hypochlorite solution temperature (5, 10, 20, 30°C), lettuce (10 g) was immersed in a sterile beaker containing 200 mL of sodium hypochlorite solution (100, 200 ppm) for 1 min and washed in 200 mL of tap water for 1 min. Lettuce (10 g) was stomached in stomacher bags containing 90 mL of 0.1% peptone water. Additional dilutions were prepared in sterile 0.1% peptone water. Appropriate dilutions were pour-plated onto tryptic soy agar (TSA, Difco, Detroit, MI), violet red bile agar (VRBA, Difco), and mannitol salt agar (MSA, Difco) and were incubated at 37°C for 24 ± 1 h. All experiments were replicated three times. Colonies were counted and the results were expressed as colony-forming unit (CFU)/g and converted to \log_{10} CFU/g values.

Results and Discussion

Optimum concentrations of sodium hypochlorite solution as a disinfectant and sanitizer. Fig. 1 shows the reduction rate of microorganisms in lettuce at various concentrations (50-500 ppm) of a sodium hypochlorite solution. The initial contamination levels of total aerobic bacteria, *E. coli*, and *S. aureus* before treatment were 2.89, 3.75, and 3.89 log CFU/g, respectively. Treatment with 50 ppm sodium hypochlorite solution reduced these levels to 0.63, 1.53, and 1.63 log CFU/g, respectively. Total aerobic bacteria were not detected in lettuce after treatment with a 100 ppm sodium hypochlorite solution. *E. coli* and *S. aureus* were not detected after treatment with 250 and 300 ppm sodium hypochlorite solutions, respectively. Park *et al.* [2004] reported that treatment with a 200 ppm sodium hypochlorite solution for 5 min failed to reduce the level of *B. subtilis* by 90%, but treatment with a 200 ppm sodium hypochlorite solution for 1 min significantly reduced the level of *L. monocytogenes*.

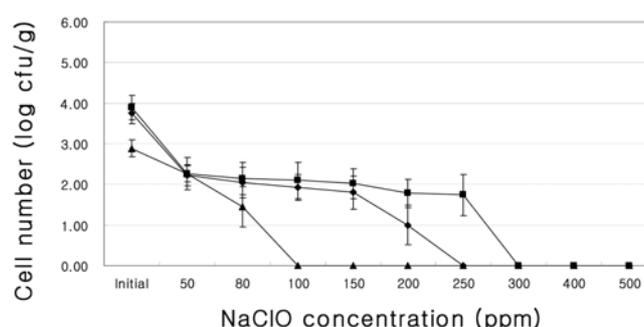


Fig. 1. Reduction rate of microorganisms in lettuce at different concentrations of a sodium hypochlorite solution. ◆, *E. coli*; ■, *S. aureus*; ▲, total aerobic bacteria.

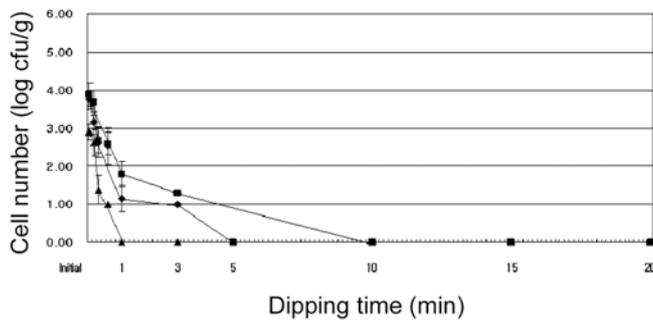


Fig. 2. Reduction rate of microorganisms in lettuce at different immersion times in a sodium hypochlorite solution (200 ppm). ◆, *E. coli*; ■, *S. aureus*; ▲, total aerobic bacteria.

Jeong *et al.* [2006] reported that initial *E. coli* contamination levels of 39 CFU/mL in lettuce were reduced to 29 CFU/mL after treatment with a 50 ppm sodium hypochlorite solution, and to 1 CFU/mL after treatment with a 100 ppm solution. The level of *S. aureus* in lettuce was reduced from 51 CFU/mL to 31 CFU/mL after treatment with a 20 ppm sodium hypochlorite solution while treatments with 50 and 100 ppm solutions reduced the levels to 39 and 6 CFU/mL, respectively. Kim [2005] found that initial contamination levels of total aerobic bacteria and coliforms in lettuce of 6.13 log CFU/g and 3.85 log CFU/g, respectively, were reduced to 3.24-4.42 log CFU/g after immersion in chlorine solutions in a commercial facility. Grag N *et al.* [1990] reported that total aerobic bacteria in lettuce was reduced from 6.04 log CFU/g to 3.47 log CFU/g after immersion in a 300 ppm chlorine solution. However, lettuce treated with highly concentrated solutions of sodium hypochlorite can suffer tissue damage, changes in taste, and a chlorine odor. These problems resulted in a 200 ppm sodium hypochlorite concentration to be set as the legal maximum permissible concentration in Korea.

Optimum immersion time. Fig. 2 shows the reduction rate of microorganisms in lettuce immersed for 20 min in a 200 ppm sodium hypochlorite solution. Levels of total aerobic bacteria, *E. coli*, and *S. aureus* in lettuce washed with tap water were reduced 0.28, 0.61, and 0.22 log CFU/g, respectively. However, the levels of microorganisms after washing with tap water subsequent to immersion in a 200 ppm sodium hypochlorite solution for 10 sec were reduced 1.5, 1.14, and 1.22 log CFU/g, respectively. Total aerobic bacteria, *E. coli*, and *S. aureus* were not detected when lettuce was immersed in a 200 ppm sodium hypochlorite solution for 1, 5, and 5 min. Cho *et al.* [2004] observed that treatment with a 100 ppm chlorine solution for 2 min reduced the initial the levels of total aerobic bacteria and *E. coli* in cabbage by 2 log CFU/g.

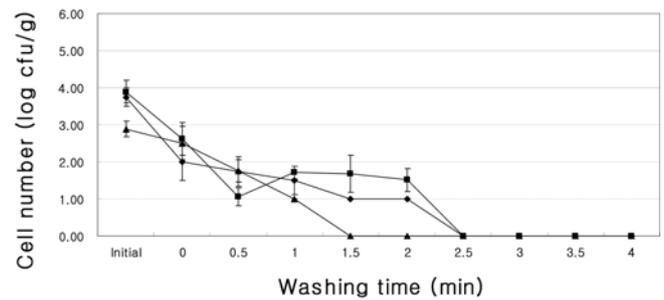


Fig. 3. Reduction rate of microorganisms in lettuce at different washing times (min) with tap water after treatment with a sodium hypochlorite solution (200 ppm). ◆, *E. coli*; ■, *S. aureus*; ▲, total aerobic bacteria.

Kim *et al.* [2004] reported that treatment with a 100 ppm chlorine solution for 3-5 min reduced levels of total aerobic bacteria and coliforms from 6.88 log CFU/g and 5.71 log CFU/g to 5.51 log CFU/g and 3.28 log CFU/g, respectively. The Korean Food Code specifies that vegetables in a lunch box product should be immersed in a 100 ppm sodium hypochlorite solution for 10 min based on the idea that initial vegetable microorganism levels of 3-4 log CFU/g will be sufficiently reduced by immersion for 1-5 min in a chlorine solution. Fig. 3 shows the reduction rate for microorganisms in lettuce with different washing times using tap water after treatment with a sodium hypochlorite solution. Total aerobic bacteria, *E. coli*, and *S. aureus* in lettuce treated with a 200 ppm sodium hypochlorite solution without rinsing were reduced 0.38, 1.74, and 1.27 log CFU/g, respectively. Washing for 30 sec after treatment with a 200 ppm sodium hypochlorite solution reduced the initial levels 1.12, 2.00, and 2.83 log CFU/g, respectively. Total aerobic bacteria, *E. coli*, and *S. aureus* were not detected when lettuce was immersed in a sodium hypochlorite for 1.5, 2.5, and 2.5 min, respectively. Kim and Kim [2005] reported that the initial contamination level of total aerobic bacteria (1×10^7 CFU/g) was proportionally reduced with an increase in the washing time. Immersion in a chlorine solution is an important process for reducing the number of microorganisms in salad. However, textural and sensory characteristics of vegetables were degraded with washing for more than 2 min.

Optimum temperature. Fig. 4 shows the reduction rate for microorganisms in lettuce at various temperatures (5, 10, 20, and 30°C) of a sodium hypochlorite solution. Temperature affected the reduction levels of total aerobic bacteria, *E. coli*, and *S. aureus* in lettuce after treatments with 100 and 200 ppm sodium hypochlorite solutions. According to Zhuang *et al.* [1995], when warm tomatoes (26 to 40°C) were immersed at 20-22°C, the reduction

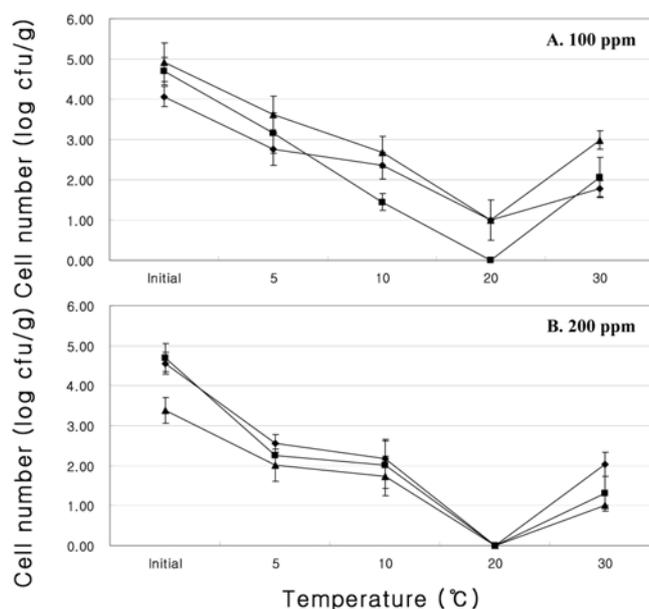


Fig. 4. Reduction rate of microorganisms in lettuce at different temperatures of a sodium hypochlorite solution. ◆, *E. coli*; ■, *S. aureus*; ▲, total aerobic bacteria.

level was decreased due to the generation of negative pressure when a vegetable is stored at a temperature lower than the temperature of the vegetable. The temperature of wash-water should be higher than the temperature of the vegetables it in order to minimize uptake of microorganisms into the vegetable tissue. We found that the levels of total aerobic bacteria, *E. coli*, and *S. aureus* in lettuce were influenced by the concentration of a sodium hypochlorite solution, the immersion time, the temperature, and the washing time. Use of optimum sanitation conditions will improve the safety and the added value of fresh-cut lettuce without affecting the sensory characteristics.

Acknowledgment. This work was supported by a grant from the Korea Food and Drug Administration in 2008 (08062sicpuman012) and the Gyeonggi Regional Research Center (20080577).

References

- Bae HJ (2001) Survey on sanitation practice and the analysis of improvement by implementing HACCP system in foodservice operation. PhD Thesis, Sookmyung University, Seoul, Korea.
- Bergdoll MS (1979) In *Foodborne Infections and Intoxication*. Bryan FL (2nd ed), pp. 12. Academic Press, New York, NY, U.S.A.
- Beuchat LR and Golden DA (1985) Antimicrobials occurring naturally in food. *Food Technol* **43**, 134-142.
- Cho JI, Kim KS, Bahk GJ, and Ha SD (2004) Microbial assessment of wild cabbage and its control. *Korean J Food Sci Technol* **36**, 162-167.
- Choi MR and Lee SY (2008) Inhibitory effects of chlorine dioxide and a commercial chlorine sanitizer against food-borne pathogens on lettuce. *Korean J Food Cookery Sci* **24**, 445-451.
- Gardial SF, Clemons DS, Woodruff RB, Schumann DE, and Bums MJ (1994) Comparing consumer's recall of prepurchase and postpurchase product evaluation experiences. *J Consumer Res* **20**, 548-560.
- Grag N, Churey KJ, and Splittstoesser DF (1990) Effect of processing conditions on the microflora of fresh-cut vegetable. *J Food Prot* **53**, 701-708.
- Jeong JW, Kim TK, Park JW, and Lee KG (2006) Antimicrobial activity of sterilizer for the exclusive of vegetable against *Escherichia coli* and *Staphylococcus aureus*. *Food Engin Progr* **10**, 221-225.
- Jung JW, Park KJ, Park BI, and Kim YH (1996) Surface sterilization effect of electrolyzed acid-water on vegetable. *Korean J Food Sci Technol* **28**, 1045-1051.
- Kim BS, Jung JW, Jo JH, and Park HW (2002) Development of surface sterilization system for fresh leafy vegetable. F02303-0252 Korea Food Research Institute Korea. pp. 25.
- Kim JM (2001) Use of chlorine dioxide as a biocide in the food industry. *Korea J Food Nutr* **6**, 33-39.
- Kim JW and Kim SH (2005) Establishment of washing conditions for salad to reduce the microbial hazard. *Korean J Food Cookery Sci* **21**, 703-708.
- Kim MH (2005) Studies on microbiocidal effect and quality preservation of fresh food by electrolyzed water. PhD Thesis, Pukyong National University, Busan, Korea.
- Kim MH, Jeong JW, and Cho YJ (2004) Cleaning and storage effect of electrolyzed water manufactured by various electrolytic diaphragm. *Korean J Food Preserv.* **11**, 160-169.
- Kim YJ, Cho JI, and Kim KS (2002) Evaluation of washing treatment using chlorine and heat hurdles on natural microflora in fresh lettuces collected from domestic markets. *Food Engin Progr* **6**, 329-335.
- Kunimoto H (1997) Acidic electrolyzed saline solution: It antimicrobial activity and factor, and practical application. MS Thesis, Korea University, Seoul, Korea.
- Manvell PM and Ackland MR (1986) Rapid detection of microbial growth in vegetable salads at chill and abuse temperature. *Food Microbiol* **3**, 59-65.
- Marshall JJ, Duxbury L, and Heslop LA (1995) Coping with household stress in the 1990s: who uses convenience foods and do they help?. *Adv Consumer Res* **22**, 729-734.
- No SY (2003) Effect of lamp type ozone generator on inactivation of microorganisms and product quality of Angelica keiskei, PhD Thesis, Yonsei University, Seoul, Korea.
- Park HK and Kim SB (2004) Microbial reduction of fresh vegetable by treatment of sanitizing reagents. *Korean J*

- Food Nutr* **17**, 436-441.
- Peter R, Wim V, Frank D, and Johan D (2004) Consumer perception and choice minimally processed vegetables and packaged fruits. *Food Qual Prefer* **15**, 259-270.
- Riley LW, Remis RS, Helgerson DD, and Mcgee HB (1983) Hemorrhagic colitis associated with a rare *Escherichia coli* serotype. *New Engl J Med* **308**, 681-685.
- Sakai S (1995) Application and development of electrolyzed oxidizing water. *Food Ind* **4**, 35-41.
- Takeuchi K, and Frank JF (2000) Penetration of *E. coli* 0157:H7 into lettuce tissue as affected by inoculum size and temperature and the effect of chlorine treatment on cell viability. *J Food Protect* **63**, 434-440.
- U.S Food and Drug Administration. (2008) <http://www.fda.gov>, The outbreak profile of foodborne
- Zhuang RY, Beuchat LR, and Angulo FJ (1995) Fate of *Salmonella Montevideo* on and in raw tomatoes as affected by temperature and treatment with chlorine. *Appl Environ Microbiol* **61**, 2127-213.