The Chemopreventive Potential and Anti-inflammatory Activities of Korean Black Ginseng in Colon26-M3.1 Carcinoma Cells and Macrophages

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We investigated the chemopreventive and anti-inflammatory activity of Korean black ginseng (BG) in colon26-M3.1 carcinoma cells and macrophages (*in vitro*) compared with white ginseng (WG). Both WG and BG had no significant inhibitory effect on normal splenocytes, but on colon26-M3.1 carcinoma cells, BG showed significantly greater inhibitory effect than that of WG. Treating with either WG or BG extracts, the release of TNF- α was significantly decreased in response to lipopolysaccharide as compared with untreated control.

Key words: black ginseng, colon26-M3.1 carcinoma cells, cytokines, white ginseng

Today basic science and therapy based clinical trials are actively exploring the potential of Complementary and Alternative Medicine (CAM), which has been rapidly accelerated with the commencement of the National Center for CAM. Studies of the immune enhancing properties of CAM based therapies have demonstrated the importance of CAM therapies in modulating the immune systems [Manardi *et al.*, 2009]. The current study has uncovered the biochemical mechanism involved in immuno modulatory pathways triggered by three types of ginseng. Several ongoing clinical trials are investigating the potential benefits of CAM based immune modulating therapies for asthma, allergic rhinitis, and atopic dermatitis [Gillis, 1997; O'Hara *et al.*, 1998;

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Manardi *et al.*, 2009]. Ginseng radix (*Panax ginseng* C.A. Meyer, *Araliaceae*) is one of the most widely used natural immuno modulatory resources in Asian.

Among Korea ginsengs, white ginseng (WG) is dried root of ginseng, and BG is the root of ginseng which is steamed at about 100°C and dried by 9 cycles of steam treatment, which makes it black in color [Lee et al., 2006]. Black ginseng (BG) getting through steam treatment had showed the enhanced pharmacological activities resulting from the enrichment chemical constituents, including ginsenosides. The ginsenosides have been focused currently on their bioactive properties [Keum et al., 2003]. It has been reported that main ginsenosides of BG are ginsenosides Rg₃ Rh₁ or Rh₂ [Kim et al., 2000; Nam, 2005] which can reduce the proliferation of various type of cultured cells [Kim et al., 1996; Kim et al., 2000; Lee et al., 2006]. The pharmacological and biological activities of steamprocessed ginseng are greater than non-steamed ginseng. During the steaming process, the ratio of major bioactive

components, including ginsenoside saponins, phenolics, and protein, is altered by newly produced components [Hasegawa, 1997a; Yun, 2003] with anti-carcinogenic, anti-stress, and antioxidant effects in humans and animals [Kang *et al.*, 2006].

In this current study, we compared BG, enriched ginsenosides including Rg_{3} , Rh_1 and Rh_2 compared with WG, to evaluate the chemopreventive and anti-inflammatory activities using *in vitro* models.

The two types of Korean ginseng, WG and BG, were donated by Bomun P & F. Ltd. (Keumsan, Korea) and identified by Dr. Kang, Department of Oriental Medicine Resources, Joongbu University (Keumsan, Korea). All two types of ginseng were extracted three times with 70% (v/v) ethanol at room temperature for 1 h and the compositions of the extracts were analyzed using LC/MS methods for ginsenosides. Then, the extracts were collected and concentrated to 65°brix. The concentrated extracts were stored in a -70° C cryogenic freezer (NU-6518G, NuAire, Plymouth, NY) and used as experimental samples.

Six to seven week old female specific pathogen-free BALB/c female mice were purchased from NARA biotech.(Seoul, Korea) and maintained at the Laboratory of Animal Experiments, Department Food & Nutrition, Yuhan University, Korea. Experiments were conducted in accordance with the guidelines established by the Animal Care and Use Committee of Yuhan University (2009E-001). Water and commercial pellet diets were supplied *ab libitum*.

A highly metastatic cell line of Colon 26 carcinoma (colon26-M3.1) were maintained as monolayer cultures in EMEM supplemented with 7.5% fetal bovine serum, sodium pyruvate, nonessential amino acids and L-glutamine. Splenocytes from mice were cultured in RPMI-1640 (Gibco, Carlsbad, CA) supplemented with 7.5% FBS [Yoon *et al.*, 1995].

Murine tumor cells and normal splenocytes from mouse in 96 well plates were incubated with various doses of the two types of Korean ginseng (5000-40 mg/ mL) for 72 h. Cytotoxicity against tumor cells and splenocyte was assayed by a cell counting kit (Wako Pure Chemical Industries, Ltd., Osaka, Japan). The absorbance of each well was monitored at 450 nm using a microtiter plate reader (Molecular Device, Sunnyvale, CA).

Peritoneal macrophages were harvested from thioglycollatetreated mice as described previously [Ha *et al.*, 2004]. Cells $(1 \times 10^{6}/mL/well)$ suspended in complete RPMI-1640 medium were plated into 24 well culture plates. After 2 h of incubation, non-adherent cells were removed by washing with PBS, the adherent macrophages were co-incubated with various doses (0-200 µg/mL) of two types of Korean ginseng extracts and/or lipopolysaccharide (LPS) (1 μ g/mL) for 24 h. The concentration of TNF- α in the culture supernatants was determined using ELISA kits (Pharm-in-gen, San Jose, CA) in accordance with the manufacturer's recommendations.

The means and standard deviations were calculated for all experiments. The data were analyzed by the analysis variance procedure of SAS (SAS analysis system, ver. 8.1). The significance of differences between groups was determined using Student's t-test.

It has been reported that ginsenosides and their metabolites mainly show the pharmacological activities of ginseng [Yun, 2003; Attele et al., 1998]. Ginsenosides are diverse group of triterpenoidal saponines showing the ability to target a vast range of tissues. The two major structural groups of ginsenosides are panaxadiols and panaxtriols. The panaxadiol group includes Rb1, Rb2, Rc, Rd, Rg3, Rh2, and Rh3, and panaxtriol group includes Re, Rf, Rg1, Rg2, and Rh1. Among these ginsenosides, Rh2 and Rg3 are main active components in ginseng radix. It has been reported that Compound Rh2 reduced the proliferation of various cultured cancer cells, and can influence apoptosis [Fei et al., 2002; Popovich and Kitts, 2002]. Rg3 has also been shown to possess anti-tumor properties, and has an effect on drug-resistant, cultured cancer cells [Keum et al., 2003; Kim et al., 2003]. Based on our preliminary study, by 9 times steaming, the level of Rg3 was significantly enriched. The levels of Rg₃, Rh₁ and Rh₂, which have anti-tumor effects [Kim et al., 2000; Nam, 2005], in the WG and BG extracts were shown in Table 1. The level of Rg₃ in BG was significantly greater than that of WG. In this study, we report that ginsenoside Rg₃ and Rh₁ or Rh₂ of BG has been enriched during the steaming process. But, there were no Rg₃. Rh₁ and Rh₂ in WG; this result is very interesting since they have the ability to reduce the proliferation of various cultured cells. This result is consistent with suggestions that the pharmacological and biological activities of steam ginseng are greater than non-steamed ginseng [Hasegawa et al., 1997b; Yun, 2003].

Table 1. Ginsenoside contents of the WG and BG (unit: mg/mL)

Analytical items	White ginseng	Black ginseng
ginsenoside Rg ₃	$ND^{1)}$	4.162 ± 0.010^{2} *
ginsenoside Rh	ND	0.032 ± 0.001
ginsenoside Rh_2	ND	0.726 ± 0.016

¹⁾Non-detectable

²⁾Values are mean±SD (n=3). The Student's t-test was done to perform a statistical comparison between WG extract and BG extract at *p<0.05.



Fig.1. Effect of samples on normal splenocytes. (\blacksquare), white ginseng extract; (\bigcirc), black ginseng extract. Data were expressed as mean value±SD (n=3). *p<0.05.



Fig. 2. Cytotocixity of samples on colon26-M3.1 carcinoma cells. (\blacksquare), white ginseng extract; (\bigcirc), black ginseng extract. Data were expressed as mean value±SD (n=3). *p<0.05.

The cytotoxic effects of WG and BG extracts in normal splenocytes and colon26-M3.1 carcinoma cells (in vitro) are shown in Fig. 1 and Fig. 2. None of the extracts showed any significant inhibitory effect on normal splenocytes at a concentration which is less than 1250 μ g/ mL, but on colon26-M3.1 carcinoma cells, BG showed significantly greater inhibitory effects comparing with that of WG. The IC₅₀ values in colon26-M3.1 carcinoma cells were 2000 µg/mL (WG), and 800 µg/mL (BG), respectively. In other word, the effect of BG was more than 2 times stronger than that of WG. Thus BG had more proliferation preventive effect on colon26-M3.1 carcinoma cells. The greater anti-proliferative effect of BG in colon26-M3.1 carcinoma cells may be due to the greater amounts of Rg_3 Rh₁ and Rh₂, which were enriched during steaming process [Wargovich, 2001; Song et al., 2006].

The effects of WG and BG extracts on macrophagemediated tumor necrosis factor-alpha (TNF- α) production



Fig. 3. The effect of WG and BG on TNF- α proliferation. (\Box), white ginseng extract; (\blacksquare), black ginseng extract. The values are expressed as the mean±SD (n=3). The Student's t-test was done to perform a statistical comparison between LPS stimulated cell non-treated WG or BG extracts and treated cell with various dose of WG or BG extracts. The asterisks indicate significant differences from the LPS (1 µg/mL) stimulated cell non-treated WG or BG extracts (*p<0.05 and ** p<0.01).

is shown in Fig. 3. After treatment with either WG or BG extracts, significantly, and dose-dependently, decreasing the releasing of TNF- α at concentrations up to 200 or 100 µg/mL, respectively, in response to LPS as compared with WG treated group. LPS, which is the component of out membrane of gram-negative bacteria and has been used to evaluate the anti-inflammatory or immunostimulatory effect of various materials [Chun et al., 2007]. LPS-activated macrophages induced the production of proinflammatory cytokines such as TNF- α , IL-1 β and IL-6 [Delgado *et al.*, 2003]. Among these cytokines, TNF- α is a potent cytokine acting beneficial functions in the activation and regulation of the host defense system, as well as inflammatory and cytotoxic activities of the host [Croft, 2009]. TNF- α produced from macrophage acts in both non-specific defense (or innate immunity) as well as to help initiate specific defense mechanisms (or cellmediated immunity) of vertebrate animals. TNF- α has been reported that the important cytokine to activate T cells and to reject tumor cells in antitumor mechanism, and has been extensively tested in vitro and in vivo, as well as in clinical trials for immunotherapy of malignant diseases [Tanigawa et al., 2000]. In addition, TNF- α has been implicated as an important mediator of cardiovascular complications such as acute myocardial infarction [Maury and Teppo, 1989; Hasegawa et al., 1997a], ischemia-reperfusion injury [Squadrito et al., 1993; Gurevitch et al., 1996; Gurevitch et al., 1997], atherosclerosis [Barath et al., 1990], chronic failure [Levine et al., 1990; Fichtlscherer et al., 2001] and

coronary artery disease [Seiler *et al.*, 2003]. Thus, the present study supports that BG has anti-inflammatory effect on the production of TNF- α in LPS activated macrophages, and alters susceptibility to microbial infections and inflammatory diseases.

In conclusion, we report that BG extract exerts anticarcinogenic effects on colon26-M3.1 carcinoma cells, and does not participate in the cytokine production (TNF- α) induced by in response to LPS as compared with untreated controls on the marcrophage-mediated cell line. Further research is needed to elucidate the effects of BG, defining the anti-inflammatory activities relationship of ginsenosides *in vivo* study.

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References

- Attele AS, Wu JA, and Yuan CS (1998) Ginseng pharmacology: multiple constituents and multiple actions *Biochem Pharmaco* 58, 1685-1693.
- Barath P, Fishbein MC, Cao J, Berenson J, Helfant RH, and Forrester JS (1990) Detection and localization of tumor necrosis factor in human atheroma *Am J Cardiol* 65, 297-302.
- Chun SC, Jee SY, Lee SG, Park SJ, Lee JR, and Kim SC (2007) Anti-inflammatory activity of the methanol extract of Moutan cortex in LPS-activated Raw264.7 cells. *Evid Based Complement Alternat Med* **4**, 327-333
- Croft M (2009) The role of TNF superfamily members in Tcell function and diseases. *Nat Rev Immunol* 9, 271-285
- Delgado AV, McManus AT, and Chambers JP. (2003) Production of tumor necrosis factor-alpha, interleukin 1-beta, interleukin 2, and interleukin 6 by rat leukocyte subpopulations after exposure to substance P. *Neuropeptides* **37**, 355–361.
- Fei XF, Wang BX, Tashiro S, Li TJ, Ma JS, and Ikejima T (2002) Apoptotic effects of ginsenoside Rh2 on human malignant melanoma A375-S2 cells. *Acta Pharmacol Sin* 23, 315-322.
- Fichtlscherer S, Rossig L, Breuer S, Vasa M, Dimmeler S, and Zeiher AM (2001) Tumor necrosis factor antagonism with etanercept improves systemic endothelial vasoreactivity in patients with advanced heart failure. *Circulation* **104**, 3023-3025.
- Gillis CN (1997) Panax ginseng pharmacology: A nitric oxide link? *Biochem Pharmacol* 54, 1-8.
- Gurevitch J, Frolkis I, Yuhas Y, Paz Y, Matsa M, and Mohr R (1996) Tumor necrosis factor-alpha is released from the isolated heart undergoing ischemia and reperfusion. *J*

Am Coll Cardiol 28, 247-252.

- Gurevitch J, Yuhas Y, Lifschitz-Mercer B, Berger E, and Paz Y (1997) Anti-tumor necrosis factor-alpha improves myocardial recovery after ischemia and reperfusion. J Am Coll Cardiol 30, 1544-1561.
- Ha ES, Hwang SH, Shin KS, Yu KW, Lee KH, Choi JS, Park WM, and Yoon TJ (2004) Anti-metastatic activity of glycoprotein fractionated from Acanthopanax senticosus, involvement of NK-cell and macrophage activation. *Arch Pharm Res* **27**, 217-224.
- Hasegawa H, Sung JH, and Huh JD (1997a) Ginseng intestinal bacteria metabolite IH901 as a new antimetastatic agent. *Arch Pharm Res* **20**, 539-544.
- Hasegawa H, Sung JH, Matsumiya S, and Uchiyama M (1997b) Main ginsengsaponine metabolites formed by intestinal *Prevotella oris* in hydrolyzing saponines. *Plant Med* 63, 436-440.
- Kang KS, Kim HY, Pyo JS, and Yokozawa T (2006) Increase in the free radical scavenging activity of ginseng by heat-processing. Biol Pharm Bull 29, 750-754.
- Keum YS, Han SS, Chun KS, Park KK, Park JH, and Lee SK (2003) Inhibitory effects of the ginsenoside Rg3 on phorbol ester-induced cyclooxygenase-2 expression, NFkappaB activation and tumor promotion. *Mutat Res* 523-524, 75-85.
- Kim SI, Park JH, Ryu JH, Park JD, Lee YH, and Park JH(1996) Ginsenoside Rg5, a genuine dammarane glycoside from Korean red ginseng. *Arch Pharm Res* 19, 551-553.
- Kim SW, Kwon HY, Chi DW, Shim JH, Park JD, and Lee YH (2003) Reversal of P-glycoprotein-mediated multidrug resistance by ginsenoside Rg3. *Biochem Pharma*col 65, 75-82.
- Kim WY, Kim JM, Han SB, Lee SK, Kim ND, and Park MK (2000) Steaming of ginseng at high temperature enhances biological activity. J Nat Prod 63, 1702-1704.
- Lee JH, Shen GN, Kim EK, Shin JH, Myung CS, Oh HJ, Kim DH, Roh SS, Cho W, Seo YB, Park YJ, Kang CW, and Song GY (2006) Preparation of black ginseng and its antitumor activity. *Korean J Orient Physiol Pathol* 20, 951-956.
- Levine B, Kalman J, Mayer L, Fillit H, and Packer M (1990) Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. N Engl J Med 323, 236-241.
- Manardi T, Kapoor S, and Bielory L (2009) Complementary and alternative medicine: herbs, phytochemicals and vitamins and their immunological effects. J Allergy Clin Immunol 123, 286-295.
- Maury CP and Teppo AM (1989) Circulating tumour necrosis factor-alpha (cachectin) in myocardial infarction. *J Int Med* **225**, 333-336.
- Nam KY (2005) The comparative understanding between red ginseng and white ginseng. *J Ginseng Research* 29, 1-18.
- O'Hara M, Kiefer D, Farrell K, and Kemper K (1998) A

review of 12 commonly used medicinal herbs. *Arch Fam Med* 7, 523-536.

- Popovich DG and Kitts DD (2002) Structure-function relationship exists for ginsenosides in reducing cell proliferation and inducing apoptosis in the human leukemia (THP-1) cell line. *Arch Biochem Biophys* **406**, 1-8.
- Seiler C, Pohl T, Billinger M, and Meier B (2003) Tumor necrosis factor alpha concentration and collateral flow in patients with coronary artery disease and normal systolic left ventricular function. *Heart* 89, 96-97.
- Song GY, Oh HJ, Roh SS, Seo YB, Park YJ, and Myung CS (2006) Effect of black *ginseng* on body weight and lipid profiles in male rats fed normal diets. *Korean J Pharmacog* 50, 381-385.
- Squadrito F, Domenica A, Zingarelli B, Ioculano M, Calapai G, and Campo GM (1993) Tumor necrosis factor involvement in myocardial ischaemia-reperfusion injury. *Eur J Pharmacol* 237, 223-230.

- Tanigawa K, Craig RA, Stoolman LM, and Chang AE (2000) Effects of tumor necrosis factor-alpha on the in vitro maturation of tumorreactive effector T cells. J Immunother 23, 528–35.
- Wargovich MJ (2001) Colon cancer chemoprevention with ginseng and other botanicals. *J Korean Med Sci* 16, S81-86.
- Yoon TJ, Yoo YC, Choi OB, Do MS, Kang TB, Lee SW, Azuma I, and Kim JB (1995) Inhibitory effect of Korean mistletoe (*Viscum album* coloratum) extract on tumour angiogenesis and metastasis of haematogenous and nonhaematogenous tumour cells in mice. *Cancer Lett* 20, 83-91.
- Yun TK (2003) Experimental and epidemiological evidence on non-organ specific cancer preventive effect of Korean ginseng and identification of active compounds. *Mutat* Res 523, 63-74.