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An in-depth study on noncovalent stacking interactions between DNA bases and aromatic drug fragments using DFT method and AIM analysis: conformers, binding energies, and charge transfer

Hossein Azizi Toupkanloo¹ · Zoha Rahmani²

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Abstract This work is aimed at providing physical insights about the π - π stacking interactions of some popular drug fragments (DF) including indole (I), benzothiophene (Bt), benzofuran (Bf) and guanine (G), adenine (A), A-thymine (AT), G-cytosine (GC) base pairs using density functional theory (DFT), the atoms in molecule (AIM) theory, and natural bond orbital (NBO) analysis. Several stable conformers of present molecules and complexes were optimized at the M062X/6-311++G(d,p) level of theory. The result shows that the IG1 (see the notation below) and IA6 have maximum interaction energy in all of the two G-based and A-based conformers; and order of the adsorption strength is IG1 > BtG6 > BfG1for G-based complexes and IA6 > BtA6 > BfG6 for A-based complexes. For the base pair-drug fragment complexes, the order of interaction energy was found according to IAT4 > BtAT3 > BfAT4 and IGC3 > BtGC2 > BfGC2, for AT and GC base pairs,

Interaction energy: This is the contribution to the total energy that is caused by an interaction between the objects being considered. π - π stacking: This parameter refers to attractive, noncovalent interactions between aromatic rings, since they contain π -bonds, namely the face-centered stacked arrangement.

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Hossein Azizi Toupkanloo h.azizi@neyshabur.ac.ir

- ¹ Department of Physics and Chemistry, Faculty of Science, University of Neyshabur, P. O. Box 9319774400, Neyshabur, Iran
- ² Computational Quantum Chemistry Laboratory, Department of Chemistry, University of Sistan and Baluchestan, P.O. Box 98135-674, Zahedan, Iran

respectively. Furthermore, our results show that stacking interaction leads to an increase and decrease in hydrogen bond length that involved in the nucleic base–drug fragment interactions. DFT-calculated interaction energies for all present conformers were found to be in a good agreement with the bond critical points data from AIM analysis. In contrast, no reasonable linear correlation was observed between NBO analysis and stability of the all studied conformers. Finally, in order to verify the DFT and AIM results, docking calculations were performed using AutoDock software. According to the binding energy of drug–DNA from AutoDock calculations, the D2-Bt and D1-Bf are the most and the least stable structures, respectively.

Keywords AutoDock \cdot Binding energy \cdot Drug fragment \cdot Interaction energy $\cdot \pi - \pi$ stacking

Introduction

Noncovalent interactions play a unique role in biological science, control diverse phenomena including boiling points of liquids, solvation energies and determine the structures of DNA, RNA, and proteins. The π -interactions including π -stacking, cation- π , anion- π , and X–H··· π constitute one of the most important chemical classes of noncovalent interactions that contribute to biomolecular structure, drug binding, and the structure and properties of π -conjugated materials of interest in organic electronics and photonics [1–5]. Among the most prevalent noncovalent interactions, the preferred orientation of the aromatic groups may be related to the π - π interactions [6]. To illustrate the importance of the issue, the aromatic–aromatic interactions play a crucial role not only in the

stabilization of proteins, but also in the recognition processes of protein-protein and protein-ligand [7]. More than half a century, researcher has been interesting to the interactions between π -systems issue [8, 9]. The results show that dispersion contributes significantly to the interaction energy (IE) is complemented by a varying degree of electrostatic interactions, where the electrostatic properties of these systems are a key determinant for their orientational preferences [10]. However, some important questions still remain regarding various classes of noncovalent interactions, such as how strong they are and how they are affected by introduction of substituents or modification with changing geometry. Therefore, in order to appropriate drug design, predict their physical properties or even choose one suitable for a specific application, it is necessary to make a link between the fundamental properties of the system such as electronic, molecular structure, as well as specific physical and chemical properties. By considering this, thorough understanding of noncovalent forces and their effects on biomolecular structure and function would aid the biotechnology and nanotechnology is possible [11]. The nucleic acids, one of the most important biological group including π -stacking interactions, have a key role in biological phenomena, because they interact reversibly with a broad range of chemical species that include water, metal ions and their complexes, small organic molecules, and proteins. Therefore, its interactions need to be studied in detail because of the importance of stacking between nucleic acid bases and others species in determining the structure and flexibility of DNA and RNA [12]. This interaction is very crucial in intercalation of drugs into DNA [13, 14]. Despite the presence of numerous published papers, there are still many questions about the characterization of interaction of drugs with nucleic acids. In this context, detailed knowledge of stacking interactions may be useful in the design of drugs intercalation within DNA [13]. Biologically, in some cases, hydrogen bond has an important role in the DNA and RNA bases. In order to this, Brovarets and coworkers [15] theoretically investigated the role of the intermolecular CH···O/N hydrogen bonds in pairs of the DNA and RNA modified bases. They established 34 CH…O and 7 CH…N interactions that were detected by quantum chemical calculations in the 39 biologically important pairs involving modified nucleobases. Stacking interactions are also important for understanding of DNA sequence-dependent polymorphism, which is mainly determined by the properties of individual base pairs [16]. The π -stacking interactions are fundamental for various phenomena such as vertical base-base interactions stabilizing the double helical structure of DNA, intercalations of drugs with DNA, the packing of aromatic molecules in crystals, the formation of the 3D nucleic acid architectures, and the tertiary structures of proteins [17–21]. So, detailed suitable information of the physical factors governing this interaction will give deeper insight into the structural and functional implications of aromatic interactions in biomolecules, particularly the nucleic acids, and to the design of new intercalating drugs with potential therapeutic value. Due to their importance, there numerous studies have been performed on the stacking interactions, both experimental [22, 23] and theoretical [24, 25]. The computational studies range from analyzing canonical base stacking [26] to unusual base stacking [27]. Among natural products which are biologically active, indoles are the most plentiful heterocycles, pharmaceuticals, agrochemicals and also are pertinent substructures in functional materials (Fig. 1). Also, the side chain of tryptophan amino acid is normally modeled with indole. Indole is composed of a benzene ring and pyrrole ring sharing one double bond. Hence, indole is an important heterocyclic system since it is often considered as a model for the chromophore of the tryptophan residue in proteins and it is the basis of drugs such as indomethacin and eletriptan [28, 29]. Other biologically active natural products are benzofuran and benzothiophene heterocyclic compounds in which а benzene ring is fused to that of a furan and thiophene molecules, respectively (Fig. 1). Benzofuran derivatives depict some potent biological properties such as antihyperglycemic, analgesic, antiparasitic, antimicrobial, antitumor, and kinase inhibitor activities [30]. The benzothiophene ring system and its derivatives are present in several drug candidates, which exhibit some interesting biological properties, for example, as antitumor [31] and antimicrobial [32]. For instance, Badichi Akher and coworkers [33] characterized the π -stacking interactions between aromatic amino acids and quercetagetin, which is one of the most representative flavonol compounds with biological and chemical activities. Their results show that the sum of donor-acceptor interaction energy between amino acids/ quercetagetin and the sum of electron densities ρ calculated at bond critical points (BCPs) between them can be useful descriptors for prediction of the interaction energy values of the complexes. Heteroaromatic rings are key components in the most known drug molecules, and they play a crucial role in molecular recognition by proteins [5]. Their unique electronic structure with a distinct π -cloud located parallel above and below the ring plane allows for a variety of interaction patterns [34]. Interaction patterns exhibited by aromatic heterocycles comprise hydrophobic, polar, hydrogen bonding (H-bonding), cation- π [35], amid- π [36], halogen- π [37], and π stacking interactions [38]. These heterocyclic compounds demonstrate diverse pharmacological properties, and flexibility in their structure means a high degree of differentiation that has been proven to be useful for the search of novel therapeutic agents. These bicyclic fragments are



Fig. 1 Structures of studied molecules

found in many drugs, and it is possible they have genotoxic effects [39]. The aim of the present work is to focus on the $\pi - \pi$ stacking interactions between some popular drug fragments (DF) including indole (I), benzofuran (Bf), benzothiophene (Bt), and natural nucleo bases (see Fig. 1) using quantum mechanical method. Several conformations were considered for each base...DF complex with respect to different molecular interactions between two units. These molecular interactions and conformational changes influence the general behavior of the compounds. Thus, the conformations and interactions provide valuable information about the favorable properties which are useful todesign a drug with higher efficacy for a desired application. Manifestation of these interactions is discussed in detail. This study can help designing new and more effective drugs on interaction with DNA.

Computational methods

Geometry optimization and vibrational frequency calculations of molecules and complexes were performed using Gaussian 09 program package at the M062X/6-311++G(d,p) level of theory [40]. The CG and AT are labeled as Cytosine...Guanine and Adenine...Thymine base pairs, respectively. All calculated IEs (kcal mol⁻¹) were performed in the gas phase. Binding energies (IEs or ΔE) of the complexes have been corrected for the basis set superposition error using counterpoise method given by Boys and Bernardi [41]. The ΔE between the DF and the DNA bases was calculated according to the following equation:

$$\Delta E = (E_{\rm Complex}) - (E_{\rm DF} + E_{\rm B}).$$

where E_{DF} and E_{B} are the energies of the drug fragments and DNA base, respectively. E_{Complex} is the energy of formed complexes between DF and DNA bases. Need to explain that DF are drug fragments including indole (I), benzothiophene (Bt) benzofuran (Bf) and B are guanine (G), adenine (A), A-thymine (AT), G-cytosine (GC) base pairs.

We have employed the M06-2X functional of Truhlar and coworkers [42] which provides remarkably accurate predictions for a wide range of chemical systems, including nonbonded interactions [43]. For general use, the M06-2X functional exhibits good results not only for noncovalent complexes, but also for other ground- and excited-state properties [44]. The natural bond orbital (NBO) analysis was performed by the NBO 5.0 program package [45]. The topological properties of electron charge density were calculated by the atoms in molecules (AIM) method [46] using the AIM2000 program package [47] at the M062X/6-311++G(d,p) level of theory. Some of the important analysis parameters such as BCP, ChelpG, and E^2 is required to explain more. A point on inter-atomic surface where the gradient of electron density is equal to zero $[\nabla \rho(\mathbf{r}) = 0]$ is defined as BCP. In the CHELPG (= CHarges from ELectrostatic Potentials using a Gridbased method) scheme by Breneman and Wiberg [48], atomic charges are fitted to reproduce the molecular electrostatic potential at a number of points around the molecule. Finally, E^2 is defined as energy value required for charge transfer from donor orbital to acceptor orbital in NBO analysis or defined as second-order interaction energy between donor and acceptor orbitals.

Guanine complexes

In order to study the interaction between guanine base and medical fragments, different conformers of Bt…G (BtG), and Bf…G (BfG), I…G (IG) complexes were optimized at the M062X/6311++G(d,p) theoretical level. Moreover,

AIM and NBO analyses were also performed on the optimized complexes at the mentioned level.

Indole-guanine (IG) complexes

Based upon distribution of IEs, seven conformers were designated (IGx; x = 1-7; see Fig. 2) which can be categorized into three groups according to the energy ranges.



Fig. 2 Conformers designated for IGx complexes

The first category included IG1 and IG4 conformers. Complex IG1 is the most stable conformer among the seven conformers. According to AIM analysis, the stability can be attributed to three BCPs between atoms in this conformation: (a) between the carbon of carbonyl in guanine base and one of the carbons of the indole cycle $O=C\cdots C$, (b) between one of the carbons of guanine base and intercycle nitrogen of indole HN…C, and (c) between one of the hydrogens of NH₂ group of guanine and the carbon of indole. In the IG4 complex, a BCP is observed between the oxygen atom in the carbonyl group of guanine and the hydrogen atom in the NH group of indole C=O…HN; high distance between the O and H atoms (2.235 Å) and unsuitable direction make it so week that its stability becomes lower than IG1 (Fig. 2). The calculated $\Sigma \rho_{\rm BCP}$ values for the intermolecular BCPs of the IG1 and IG4 complexes are in agreement with the stability of these complexes, $\Sigma \rho_{\rm BCP}$ (IG1) > $\Sigma \rho_{\rm BCP}$ (IG4). Relative orientation of two units is more appropriate for π - π stacking in conformation IG1, so due to stronger $\pi - \pi$ stacking interaction, it is more stable than the structure of IG4; guanine bends on the surface of indole molecule and decreases the π - π stacking interaction between aromatic rings. According to the IEs, the IG2 and IG5 conformations have the second order. In both structures, no BCP was observed between C=O...HN and HNH...C pairs, so those are less stable in comparison with these of the first group. Due to the O=C…C and NC…NH interactions, the IG4 conformer is more stable than IG5. In the conformer IG5, the oxygen atom of guanine interacts with the nitrogen atom of indole C=O...NH; so, this conformation is less stable due to the repulsion between the lone pairs of oxygen and nitrogen. In the IG5 conformer, guanine bends on the indole ring and the distance between the oxygen atom of carbonyl group and the hydrogen atom of indole decreases to 2.423 Å. Thus, there is a weak interaction between oxygen and hydrogen which does not exist in IG2; it makes the sum of $\rho_{\rm BCP}$ in IG5 higher than that of IG2, which is not compatible with the stabilization energy of these two conformers. According to the IE ranges, the IG3, IG6, and IG7

conformers are in the third position. Due to that no BCP between C=O…HN and HNH…C pairs was observed in all these three structures; therefore, concluded these conformers are less stable than the first group. From one point of view, in the structures of IG6 and IG3, a BCP between NC…NH pair makes IG6 and IG3 more stable than IG7. From another point of view, in the structure of IG6, there is a BCP between NC…NH pair in IG6 which was not observed in IG3 conformer, which makes IG6 more stable than IG3. None of these interactions were detected in the IG7 conformer, so it is the less stable conformer. In addition, the $\Sigma \rho_{\rm BCP}$ value is higher for IG6 conformer in comparison with two other conformers which are in excellent agreement with the calculated IEs. The ρ values calculated in the intermolecular BCPs of effective interactions in the stability of various conformers of indoleguanine complex are reported in Table 1.

Benzothiophene-guanine (BtG) complexes

According to the IE range of BtG complexes, eight conformers (BtGx; x = 1-8; see Fig. 3) can be classified in three groups. The BtG5 and BtG6 conformers are in the first category. Due to the formation of H NH ... S H-bond with a high ρ value at the BCP, BtG6 is the most stable structure. The O=C…C interaction, which is confirmed by a BCP in AIM analysis, is contributive to the stability of them in both structures. BtG1, BtG2, and BtG8 conformers make the second group. In the BtG1 and BtG2 conformers, the repulsion between lone pair electrons of nitrogen and sulfur, which is confirmed by a BCP value, decreases the stability of this conformer, but in the BtG8 conformer, the NC…S interaction is weaker due to the lower electron density on the carbon connected to the nitrogen, with a lower tendency interaction with lone pair electrons of sulfur. The BtG3, BtG4, and BtG7 conformers were established the third group. Due to the O=C…C interaction, which is confirmed by the AIM analysis, BtG4 is more stable than other two complexes. The ρ values calculated at the BCPs of the

Table 1 Stabilization energy (in kcal mol⁻¹) and the ρ values (in au) calculated in the BCPs of the effective interactions in the stability of indole–guanine complexes

	ΔE (kcal mol ⁻¹)	[C=O…HN]	[C=O…N]	[O=C…C]	[NC…NH]	[HNH…C]	$[H_2N{\cdots}C]$	$\Sigma \rho \Box 10^2$
IG1	- 15.769			9.5403	8.9996	7.1414		2.5681
IG2	- 13.704			8.2989	8.4198			1.6719
IG3	- 10.411			6.4028				0.6403
IG4	- 15.693	15.3706				7.7368		2.3107
IG5	- 13.414		11.7681				5.62	1.7388
IG6	- 10.418			7.4609	7.2393			1.4700
IG7	- 10.187							0.0000



Fig. 3 Conformers designated for BtGx complexes

effective interactions in the stability of BtG complexes are reported in Table 2.

Benzofuran-guanine (BfG) complexes

Corresponding to IE range of BfG, eight conformers (BfGx; x = 1-8; see Fig. 4) can be categorized into three groups.

Among the BfG1, BfG3, and BfG7 conformers, from the first category, BfG1 conformer is the most stable conformer. Due to the ρ value at the BCP of O=C···C interaction, BfG1 and BfG3 conformers are more stable than BfG7 conformer. In addition, the N···O interaction, which is confirmed by the AIM analysis, decreases the stability of BfG7 conformer. Orientation of units in BfG1 conformer is

Table 2 Stabilization energy ΔE (in kcal mol⁻¹) and the ρ values (in au) calculated at the BCPs of the effective interactions in the stability of benzothiophene–guanine complexes

	$\Delta E \ (\text{kcal mol}^{-1})$	[HNH…S]	[O=C…C]	[NC…S]	[C=O…S]	$[H_2N{\cdots}C]$	[N…S]	$[H_2N{\cdots}S]$	$\Sigma \rho \Box 10^2$
BtG1	- 11.168						0.00681		0.681
BtG2	- 11.046		0.00783337			0.0062676	0.00711		2.121
BtG3	- 8.951				0.007408				0.741
BtG4	- 10.003		0.00590312					0.004802	1.071
BtG5	- 12.169		0.00820479	0.007431		0.0072458			2.288
BtG6	- 12.444	0.0083549	0.0081833						1.654
BtG7	- 9.444								0
BtG8	- 11.263		0.00655607	0.007218					1.377

better than BfG3; this makes stronger the NH…O H-bond and $\pi - \pi$ stacking interaction in BfG1, such that the conformer BfG1 becomes more stable than BfG3 conformer. The BfG2 and BfG5 conformers formed the second group. Although the stability of BfG2 conformer is lower than the conformers of first group, the calculated $\Sigma \rho$ value at the BCPs of BfG2 is higher than those of other conformers. The third group includes the BfG4, BfG6, and BfG8 conformers. The O=C···C and H_2N ···C interactions, confirmed by the AIM analysis, make BfG4 more stable than BfG6 and BfG8. It is expected that the C…O interaction makes BfG8 conformer more stable than BfG6. The $\Sigma \rho$ values are in good agreement with the stabilization energy for these conformers. The $\Sigma \rho$ value calculated at the BCPs of the effective interactions in the BfG complexes is given in Table 3.

Comparison between groups

Among the most stable conformers of three categories, IG1 is conformer the most stable including $\Delta E = -15.769$ kcal mol⁻¹ (Table 4). This stability is due to the presence of O=C···C, NC···NH, and HNH···C interactions. The BtG6 conformer with ΔE equal to -12.44 kcal mol⁻¹ is second rank in the energy ranking. The calculated $\Sigma \rho$ value at the intermolecular BCPs of IG1, BtG6, and BfG1 conformers are equal to 0.025681, 0.0165382, and 0.0152308, respectively. The maximum and minimum values correspond to the most and least stable conformers, respectively. The trend of the different bonds length corresponds to the complex IE shows that in I…G complex, changes in the lengths of bonds constructing the effective indole and guanine cycles in $\pi - \pi$ stacking interactions for different conformers can impact abovementioned interactions. It is also observed that changes in the length of carbon-nitrogen bond (C8-N13) have a linear relationship with IE of I…G and the decrease in bond length leads to formation of most stable conformer. In contrast, changes in C2-N13 bond length have no reasonable relationship with IE. Moreover, there is a linear relationship between the changes of C27–N22 bond length and complex IE I…G as the bond length was decreases leads to formation of most stable conformer. In addition, there is a linear relationship between nitrogen–hydrogen bond length (N13–H14) and I…G complex IE, as the increase in bond length leads to formation of most stable conformer and an H-bond with guanine. Furthermore, as the conformers become stable, C27–O28 bond length increases in a way that the coexisting stability of N–H and C–O bonds confirms the formation of H-bond in all the complexes. In BtG and BfG complexes, there is no reasonable relationship between the change in bond length and complex stabilization energy.

Indole-adenine (IA) complexes

In order to investigate the interaction between adenine base and medical pieces, various conformers of complexes indole-A (IA), benzothiophene-A (BtA), and benzofuran-A (BfA) were optimized at M062X/6311++G(d,p) level. Moreover, according to the AIM and NBO analyses, the atomic charges were calculated by the ChelpG method (Fig. 5). Among the seven possible conformers of IA complexes that are labeled IAx (x = 1-7; see Fig. 5), IA6 is the most stable conformer. This stability can be attributed to strong $\pi - \pi$ stacking interaction that is specified by many BCPs between carbon atoms of adenine base and indole. In addition, ChelpG charges show that in all conformers, the charge transfer occurs from indole to adenine and the highest charge transfer corresponds to the most stable IA5 and IA6 conformers. In contrast, about the IA3 and IA4 conformers with the least stable, the minimum amount of charge transfers between units. In the N…N interaction (the nitrogen atoms of adenine) and indole which is specified via a BCP makes between these atoms, IA3 is the least stable in comparison with other conformers of these group. A reasonable relationship was observed between the calculated ChelpG charges for indole and the ΔE values for this type of complexes. The calculated



Fig. 4 Conformers designated for BfGx complexes

 ΔE and $\Sigma \rho$ values in the BCPs of effective interactions in the stability of IA complexes are given in Table 5.

Benzothiophene-adenine (BtA) complexes

Among the eight possible conformers of BtA complexes specified as BtAx (x = 1-8; see Fig. 6), BtA6 is the most stable conformer. The higher stability of this conformer can be attributed to the C···C interaction between carbon atoms of adenine base and benzothiophene, which is distinguished by an intermolecular BCP. BtA3 and BtA5 are the least stable conformers which have the least amount of calculated $\Sigma \rho$ at BCPs. The ChelpG charge data reveal that charge transfer is done from benzothiophene to adenine in all conformers with the exception of BtA3 and BtA4 (Fig. 6). The calculated ChelpG charges and the ρ values at the most important BCPs, and the $\Sigma \rho$ values of effective

Table 3 Stabilization energy ΔE (in kcal mol⁻¹) and the ρ values (in au) calculated at the BCPs of the effective interactions in the stability of benzofuran–guanine complexes

	$\Delta E \ (\text{kcal mol}^{-1})$	[C…O]	[O=C…C]	[HNH…O]	[N…O]	[N…O]	$[H_2N\cdots C]$	$\Sigma \rho \Box 10^2$
BfG1	- 11.441		0.00843972				0.0067911	1.5231
BfG2	- 10.853	0.007324	0.00799144				0.0065916	2.1907
BfG3	- 11.434		0.00842655	0.00817935				1.6606
BfG4	- 9.517		0.00775370				0.0064131	1.4167
BfG5	- 10.323							0
BfG6	- 8.478							0
BfG7	- 11.048				0.0071234	0.00726		1.4387
BfG8	- 8.543	0.008904						0.8904

Table 4 Relationship between bond length (in Å) and IE in IGx complexes

	$\Delta E \ (\text{kcal mol}^{-1})$	<i>r</i> _{C8–N13}	<i>r</i> _{C2–N13}	$r_{\rm N13-H14}$	<i>r</i> _{C27–N22}
IG1	- 15.769	1.3753	1.37446	1.00948	1.41806
IG2	- 13.704	1.37612	1.3746	1.00837	1.42437
IG3	- 10.411	1.3792	1.37617	1.00583	1.43008
IG4	- 15.693	1.37362	1.37496	1.01242	1.41568
IG5	- 13.414	1.37603	1.37514	1.01002	1.42044
IG6	- 10.418	1.38025	1.37802	1.0058	1.42491
IG7	- 10.187	1.3806	1.37443	1.00551	1.43149

interactions in the stability of benzothiophene-adenine complexes are reported in Table 6.

Benzofuran-adenine (BfA) complexes

Among the seven possible conformers of BfA complexes that are labeled as BfAx (x = 1-7; see Fig. 7), BfA6 is the most stable conformer. The higher stability of this conformer can be attributed to the C…C interaction between adenine base and benzofuran. BfA1 is the least stable conformer with the minimum $\Sigma \rho$ value at BCPs. The ChelpG charges show that charge transfer is done in all conformers, with the exception of BfA3 and BfA4 from benzofuran to adenine. The ΔE values, ChelpG charges, $\Sigma \rho$, and the ρ values calculated at the most important BCPs of effective interactions between BfA complexes are given in Table 7 and Fig. 7. The change in the C4-C5 bond length of indole is in a linear relationship with IE of IAx. The results show that the stability of complexes increases by increasing bond length. Moreover, there is a linear relationship between the changes of N26-H31 bond length of adenine base and the IE value of IAx complexes. Higher bond lengths correspond to the more stable conformers. Moreover, the presence of interaction between indole C4 atom and adenine base N26 atom makes IA6 the most stable conformer. There is a linear relationship between

C2–C3 and C4–C5 bonds length changes in benzothiophene and the IE values of the BtAx complexes, such that the elongation in mentioned bond length was occurred, and the stability of complexes increases. Moreover, there is a linear relationship between changes in the N26–C2 and N25–H30 bond lengths of adenine base and the IE values of BtAx complex, such that the more stable complexes are accompanied by longer bond lengths. Furthermore, interaction between C3 of benzothiophene and C20 of adenine base makes BtA6 the most stable conformer. There is no reasonable relationship between the bond length changes and the IE in BfAx. However, there is a reasonable relationship between the C16–C17 bond length and IEs of complexes. The BfAx complexes become more stable by the increase in the bond length.

The NBO analysis

Donor-acceptor IEs (E^2) were obtained from NBO analysis on the M062X/6311++G(d,p) level. In the guanine complexes, the most stable conformer is BtG6 with the most $\sum E^2$ (lpDF $\rightarrow \sigma *$ G or A), and the least stable conformer is BtG3 with the least $\sum E^2$, but in general, there is no reasonable correlation between the stabilization energy of complexes and $\sum E^2$ in the guanine complexes (Fig. S1). On the other hand, the $\sum E^2$ values decrease by the increase in the stability of conformers for the BtAx and BfAx complexes, where a logical linear correlation is observed between two parameters. However, there is no reasonable relationship between the $\sum E^2$ values and the stability of conformers for the IAx complexes (see Table S1).

Interactions of medical fragments with the AT and GC base pairs

In order to investigate the interaction of indole, benzothiophene, and benzofuran fragments with the AT and GC base pairs, all IAT, BtAT, BfAT, ICG, BtCG, and BfCG



Fig. 5 Conformers designated for IAx complexes

complexes were optimized at the M062X/6-31G(d,p) computational level.

The complexes of AT base pair

In order to study the interaction of indole with AT, four different conformers named IATx (x = 1-4; see Fig. S1)

were evaluated by IE and characterization of bond critical intermolecular interactions. For IAT4, which is the most stable conformer, no reasonable relationship is observed between the calculated $\Sigma \rho$ values (or the ρ_{max} value) at the intermolecular interactions by the AIM analysis and the IE values, while both H-bonds of the AT base pair become stronger (see Table S2). However, the $\Sigma \rho_{HB}$ value of two

Table	5 Calculated ΔE (i	in kcal mol ⁻¹), (ChelpG chi	arges (in au	1), and the	ρ ($\Sigma\rho$, in ε	u) values i	n the BCPs of	effective inter-	actions in the	stability of	indole-adeni	ne complexes	
	$\Delta E \; (\text{kcal mol}^{-1})$	q (ChelpG)	[C…C]	[C…C]	[C…C]	[N···C]	[N…C]	[HC…NH]	[HN···CH]	[C…NH]	[N…CH]	[HN···C]	$\Sigma ho\ \Box10^2$	$\rho_{\rm Max} \square 10^2$
IA1	- 9.3036	0.0163	0.0072	0.0071	0	0.0074	0	0	0	0	0	0.0072	2.898	0.742
IA2	-10.733	0.0296	0.0094	0.000	0	0	0	0	0.0080	0	0	0	1.75	0.94
IA3	-9.1895	0.0144	0.0066	0.0085	0	0	0.0081	0	0.0060	0	0	0	2.921	0.848
IA4	-9.1600	0.0052	0.0069	0.0082	0	0	0	0.0077	0	0	0.0063	0	2.919	0.82
IA5	-11.6397	0.0413	0.0072	0.0081	0.0086	0	0	0	0.0064	0.00716	0	0	3.753	0.862
IA6	- 11.968	0.0368	0.0093	0.0070	0.0069	0	0	0	0.0074	0	0	0	3.068	0.934
IA7	- 9.553	0.0224	0.0072	0.0080	0	0	0	0	0.0074	0	0.0067	0.0080	3.732	0.80

H-bonds of AT base pair is lower than that of other conformers (x = 1-3). With respect to the IAT4 conformer, the N8-H26 H-bond length of AT decreases in complexation, while other H-bond (H14...O27) becomes shorter in this process. Four conformers have been considered for the BtATx (x = 1-4; see Fig. S2) complexes in which the BtAT4 is the most stable conformer. The calculated $\Sigma \rho$ values at the observed BCPs between the Bt and AT units are in agreement with the IE values (see Table S3). Similar trends were observed for both the IE and $\Sigma \rho$ values. This tendency is also in agreement with the H…O bond length and corresponding ρ_{BCP} values. Although the most stable conformer has the longest N···H bond length and the lowest $\rho_{\rm HB1}, \, \rho_{\rm HB2}$ (correspond to the H-bonds of the AT base pair), the trends of these data are not in agreement with the IE of conformers. If only the IE values were considered, we concluded that the BtAT3 is the most stable conformer (more negative), but this trend in contrast to the H…O and N…H bond length changes.

Four conformers were also considered for BfATx (x = 1-4; see Fig. S3) complex based on the IE values; the BfAT4 is the most stable conformer (see Table S4). The calculated ρ value at the BCPs of AT base pair increase on complexation process was the highest value corresponds to the most stable BfAT4 conformer. The N8…H26 and H14…O27 bond lengths decrease on complexation, except about BfAT3 for H14…O27 bond, which makes BfTA4 the most stable conformer. With the exception of BfAT2, the trend in the IE values is in agreement with the $\Sigma \rho$ values calculated at the BCPs of intermolecular interactions.

The interaction of I, Bt, and Bf with GC

Herein, four different conformers of IGC complex, named as IGCx (x = 1-4; see Fig. S4), were evaluated via IE estimation and base pairs H-bond investigation (see Table S5). The HN…CO bond, which is predicted by a BCP in the results of AIM, between nitrogen atom of indole and carbon atom of guanine with the highest ρ_{BCP} value among different conformers, makes IIGC3 the most stable conformer. On the other hand, the lowest $\Sigma \rho_{\rm BCP}$ value of three H-bonds between G and C base corresponds to this conformer. The H13---O20 distance increases on complexation process with I, while the N6…H22 and O9…H25 distances decrease on complexation procedure. In general, it can be said that changes in the H-bond lengths have been in the similar direction of complex stability. Four conformers named BtGCx (x = 1-4; see Fig. 5) were observed for BtGC complex, in which BtGC2 has the most stable conformer. The BCP of HC...CO bond has the highest density value between carbon atoms of Bt and carbonyl group of G base. The lowest $\Sigma \rho_{\text{HBCP}}$ value for three H-bonds formed between base pairs corresponds



Fig. 6 Conformers designated for BtAx complexes

to this conformer (see Table S6). The H13...O20 and O9...H25 H-bonds become weaker with respect to the increase in the length for this conformer, while the reverse trend was observed for the N6...H22 H-bond. In general, the stabilized conformers are in agreement with the changes in these bond lengths.

Four conformers BfGCx (x = 1-4; see Fig. S6) have been considered for the BfGC complex, in which BfGC2 is the most stable conformer. The lowest $\Sigma \rho_{\text{HBCP}}$ and ρ_{Max} values were calculated between two units of this conformer (see Table S7). The length of all three H-bonds formed between G and C bases has been decreased in the BfGC2

	ΛE (kcal mol ⁻¹)	d (ChelnG)	[[][]	[CC]	ניייט	[NC]	INCH1	[NS]	IHCSI	[CS]	IHH1	ISNHI	IHNCHI	$\Sigma_{O} \square 10^2$	0.1 <u></u> 10 ²
		4 (output)	$\begin{bmatrix} 0 \\ 0 \end{bmatrix}$	ר כ	5						[TT \T7.TT]			- P [10	
BtA1	- 9.079	0.00693	0.0073	0.0076	0.0065	0.0073	0	0	0	0	0	0	0	2.878	0.763
BtA2	-10.662	0.02504	0.0069	0.0065	0.0079	0	0.0063	0.0073	0	0	0	0	0	3.495	0.787
BtA3	- 8.945	-0.00217	0.0069	0.0069	0	0	0.0067	0	0.0069	0	0	0	0	2.741	0.692
BtA4	- 9.587	-0.00033	0.0075	0.0069	0.0074	0	0.0068	0.0064	0	0	0	0	0	3.492	0.747
BtA5	- 8.827	0.00428	0.0069	0.0067	0.0065	0	0	0	0.007	0	0	0	0	2.712	0.699
BtA6	- 11.139	0.0291	0.0072	0.0082	0.0066	0	0	0	0	0.0073	0	0	0	2.931	0.823
BtA7	- 9.307	0.00934	0.0063	0.0082	0	0	0.0071	0	0	0	0.0048	0.0068	0	3.324	0.82
BtA8	-10.467	0.03207	0.0063	0.0076	0.0082	0	0.0062	0	0.0073	0	0	0	0.0065	4.207	0.816

complex on complexation. The following equations were used in our calculations (see Tables S8 and S9):

$$\begin{split} \Delta E_{\rm HB} &= E({\rm base \ pair}) - (E\ {\rm base1} + E\ {\rm base2}), \\ \Delta E_{\rm HB} &+ \Delta E_{\rm S} = \Delta E_{\rm Complex} \\ &= E_{\rm Complex} - (E\ {\rm base1} + E\ {\rm base2} + E\ {\rm drug}). \end{split}$$

H-bond deformation energy (HBDE):

$$\Delta\Delta E_{\text{HB}} = \Delta E_{\text{HB}}(2) - \Delta E_{\text{HB}}(1)$$

$$\Delta E_{\text{S}}(1) = E_{\text{Complex}} - (E \text{ base}1 + E \text{ base}2 + E \text{ drug})$$
Stacking deformation energy (SDE):

$$\Delta\Delta E_{\text{S}} = \Delta E_{\text{S}}(2) - \Delta E_{\text{S}}(1)$$

Complexes of I, Bt, and Bf with AT base pair

In all conformers of indole complex with AT base pair, the charge transfer is carried out from adenine and indole to thymine so that in the most stable conformer (IAT4), the highest and lowest charges were observed on thymine and indole, respectively. In addition, for the most stable conformer (IAT4), $\sum E^2$ has the lowest value. In the complex of Bt with AT base pair, for all conformers (except BtAT2), charge transfer occurs from adenine to Bt and thymine in a way that in the most stable conformer (BtAT3), the lowest charge was observed on the Bt surface. Furthermore, in the most stable conformer (BtAT3), $\sum E^2$ is lower than compared with BtAT1 and BtAT2 conformers, but in comparison with BtAT4 conformer it is higher. In the complex of Bf with AT base pair, for all conformers, charge transfer is performed from adenine to Bf and thymine in a way that in the most stable conformer (BfAT4), the lowest charge is seen on Bf, adenine, and thymine. Additionally, in the most stable conformer (BfAT4), $\sum E^2$ has the highest value (see Table S10).

Complexes of I, Bt, and Bf with GC base pair

In the complex of indole with GC base pair, for all conformers (except IGC1), charge transfer was observed from cytosine to guanine and indole in a way that in the most stable conformer (IGC3), the highest charge was detected on indole, adenine. Furthermore, in the most stable conformer (IGC3), $\sum E^2$ has the highest value. In the complex of Bt with GC base pair, for all conformers (except BtGC3 and BtGC4), charge transfer is done from cytosine to guanine and Bt, but in the most stable conformer (BtGC2), no reasonable relationship was found between data. Also, in the most stable conformer (BtGC2), $\sum E^2$ is lower than BtGC1 and BtGC3, but it is higher than BtGC4. For the most stable complex structure (BfGC2) of Bf with GC base pair, charge transfer is done from cytosine to guanine



Fig. 7 Conformers designated for BfAx complexes

and Bf in a way that the highest and the lowest charges are seen on guanine and Bf, respectively. Moreover, in the most stable conformer (BfGC2), $\sum E^2$ is lower than BfGC3 and BfGC4, but it is higher than BfGC1 (see Table S11).

Comparing IATx, BtATx, BfATx with IGCx, BtGCx, BfGCx

The calculated ρ_{Max} (BCP) values for the most stable conformer of the IGC complexes (IGC3) are higher than those for the most stable conformer of the IAT complexes

	$\Delta E \ (\text{kcal mol}^{-1})$	q (ChelpG)	[C…C]	[C…C]	[C…C]	[HN···C]	[N…C]	[N…CH]	[N…O]	[HN…CH]	[N…CH]	$\Sigma ho \ \Box 10^2$	$ ho_{ m Max} \square 10^2$
BfA1	- 8.648	0.005084	0.0073	0.0071	0	0.0070	0.0073	0	0	0	0	2.877	0.734
BfA2	- 9.126	0.014977	0.0070	0.008	0	0	0	0	0	0	0	1.498	0.797
BfA3	- 8.654	-0.003776	0.0079	0.0075	0	0	0	0.0061	0.0075	0	0.0066	3.559	0.791
BfA4	- 8.961	-0.006233	0.0074	0.0078	0.0069	0	0	0.0066	0	0	0	2.864	0.783
BfA5	- 9.609	0.031982	0.0065	0.0073	0.0081	0.0078	0	0	0.0071	0	0	3.684	0.807
BfA6	-10.07	0.024825	0.0069	0.0082	0.0077	0	0	0	0	0	0	2.279	0.816
BfA7	-9.731	0.016244	0.0065	0.0082	0	0.0081	0	0.007	0	0.0072	0	3.704	0.818

(IAT4). Apart from these data, the $\Sigma \rho_{Max}$ (BCP) values are high for the most stable conformers of GC complexes.

DNA docking calculations

Although quantum mechanical calculations are time-consuming for large structures, new computational techniques and softwares allow us to calculate some physical and chemical factors for great structures. In this work, using AutoDock [49] software and three sequences of DNA extracted from Protein Data Bank (PDB site) [50], intercalation of three drug fragments (DF) into DNA structure was investigated and binding energies and inhibition constant (KI) were obtained (see Table 8). The main reason to use docking was to predict the intercalation of DF process in the DNA site and observe its three-dimensional structure. After preparation of DF and three sequences of DNA extracted from PDB site (D1, D2, D3), dimensions of bounding box were determined in a way that medical fragment and intercalation site are incorporated within the box. After that, calculations were performed in order to obtain the best orientations and binding energies. Among all existing complexes, corresponding to binding energy of drug with DNA, the D2-Bt and D1-Bf are the most and the least stable structures, respectively (see Tables 9, 10). In all DFs, the binding energies of CG/CG sequence with each fragment are greater than two other sequences of DNA (see Fig. 8). It should be noted that investigated DFs are part of medical structures that docking calculations have been performed already on them. Therefore, by comparing the calculations on whole drug and DFs, it can be concluded that these calculations have good adaptability with each other. It was observed that Bt fragment separated from raloxifene has the highest binding energy average with biological bases and the medical fragment of Bf separated from befunolol has the lowest average binding energy with biological bases.

As a final discussion, from the results of our study, the following main points were presented for emphasis and importance use of calculations in drug design:

1. In this study, using quantum mechanical calculations, a comprehensive understanding of the effects of charge on the more stable states of drug fragment–

 Table 8 PDB codes of DNA sequences extracted from PDB site, interaction sites, and symbols used for sequences

PDB Code	Intercalation site	Symbol
1MTG	GA/TC	D1
1N37	CG/CG	D2
1X95	TG/CA	D3

Complexes	Binding energy (kcal mol ⁻¹)	kI (micro-molar)	Size of bounding box	Interaction
D1-I	- 4.05	1070	52,56,22	Stacking, H-bonding
D1-Bt	- 3.91	1370	54,56,22	Stacking
D1-Bf	- 3.84	1540	52,54,24	Stacking
D2-I	- 4.92	246.27	54,48,42	Stacking, H-bonding
D2-Bt	- 5.06	194.22	54,38,40	Stacking
D2-Bf	- 4.79	309.43	52,36,40	Stacking
D3-I	- 4.54	470.67	30,34,60	Stacking
D3-Bt	- 4.62	409.25	32,30,56	Stacking
D3-Bf	- 4.43	565.23	30,30,52	Stacking

Table 9 Values of binding energies, inhibition constants, and bounding box size

Table 10	Docking	score	values	and	mean	values	of	binding	energy
for medica	al fragme	nts [37	7]						

Drug	Medical fragments	Docking score	Mean value of binding energy
Almotriptan, Eletriptan, Naratriptan, Rizatriptan, Sumatriptan, Zolmitriptan	Ι	6.23	- 4.503
Raloxifene	Bt	7.9	- 4.53
Befunolol	Bf	5.9	- 4.353

DNA base complexes and the magnitude of the binding energies have been performed. Binding energy parameter allows us to determine if the drug to be prepared can form stable complexes with DNA bases or not. In other words, using this parameter, we can determine its stability before doing experimental work and cost on the drug design.

- 2. In addition to binding energy, by using AIM results, we can show that the strongest interaction between the drug and the DNA bases from which position are performed, and in the design of the drug by the experimental method, the mechanism has been conducted by the calculation results, so that we have the best design for the drug. Similar to our work, Badichi Akher and coworkers [33] investigated the π -stacking interactions between aromatic amino acids and quercetagetin (one of the most representative flavonol compounds with biological and chemical activities) using AIM analysis and they found that the ring C in quercetagetin is more suitable than the rings A and B for interaction.
- 3. Since the empirical study of functional groups effects on the drug fragment properties is more expensive and time-consuming, therefore using quantum mechanics calculations, AIM results, and NBO charge analysis providing more effective drugs for special applications.



Fig. 8 Interaction between DNA and three drug fragments by AutoDock software

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